ENTOMOSPORIUM LEAF SPOT OF PHOTINIA

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Introduction. Photinia, a shrub in the family Rosaceae, is widely grown in nurseries in the southern United States. Several species, Photinia serrulata Lindl., P. glabra (Thunb.) Maxim., and a hybrid, P. X fraseri Dress are grown for landscaping. Photinia X fraseri, is especially desirable because of its bright red, immature foliage and is the most popular species grown. All of these ornamentals are susceptible to leaf spotting caused by the fungus Entomosporium maculatum Lev., which has the perfect stage Fabraea maculata (Lev.) Atk. (=Diplocarpon maculatum (Atk. Jorstad) (10)). Entomosporium leaf spot of Photinia was first reported in Louisiana in 1957 (6) and has since appeared in Florida (1) and Virginia. Other ornamentals reported to be hosts include Juneberry (Amelanchier spp.), flowering quince (Chaenomeles spp.), loquat (Eriobotrya japonica (Thunb.) Lindl.), firethorn (Pyracantha coccinea Roem. var. formosana), Indian hawthorn (Rhaphiolepis indica Lindl.), and mountain ash (Sorbus sitchensis Roem.) (1, 5, 7, 9).

Symptoms. Leaf spots on Photinia are at first minute, slightly raised dots on either surface of the leaf. Older spots have a slightly depressed center with raised margins.

On the juvenile red-colored foliage of P. X fraseri, the young minute leaf spots (more or less circular) are a darker red than the surrounding healthy tissue, whereas older necrotic spots (up to 3-4 mm in diameter) have ashen grey centers and dark-purple margins surrounded by an indefinite lighter purplish halo. The centers of the necrotic spots on both surfaces are dotted with minute black specks, covered by a hyaline membrane which are the spore structures (acervuli) of the fungus (Fig. 1). The spots are discrete in light infections, but in heavy infections, they often coalesce to form large lesions and premature defoliation. Infection is mostly limited to the leaf blade, and occasionally a few spots may occur on petioles and tender young shoots.



Fig. 1. Entomosporium leaf spot of $\underline{Photinia}$ X $\underline{fraseri}$ showing spore-producing structures within leaf spots (22X).

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Disease Development. In the spring, infection first appears on new growth of the lowest branches and spreads gradually upward. Overwintering mycelia in fallen infected leaves from the previous year produce conidia which are dispersed by splashing water (4).

Evidence that physiologic races of the Entomosporium fungus exist was first presented by Stathis et al. (8). Isolates from pear, (Pyrus spp.), and quince (Cydonia oblonga Mill.), were highly virulent to pear and loquat but nonpathogenic to Photinia spp. In contrast, the isolates from Photinia spp. were highly virulent to Photinia, pear, and loquat. However, the isolates from loquat were weakly virulent to Photinia. Stathis postulated that the race on Photinia arose from the loquat by mutation. No evidence has been presented concerning the relationship of isolates from Rhaphiolepis sp. to isolates from other hosts.

Control. Fallen leaves should be collected in the winter before the new growth appears in the spring to reduce sources of the fungus. Plants pruned in late summer may suffer infections on resultant new growth into the fall.

Fungicides applied to pear in Georgia after the first leaves are fully expanded gave good protection (2). On quince there were more infections in a wet spring and summer than during a dry season (11). Under normal nursery production in Florida, sprinkler irrigation is practiced and this provides adequate moisture for disease development in container-grown Photinia spp. Therefore, more frequent application of fungicides will probably be necessary than under nonsprinkler irrigation systems. The fungicide benomyl (Benlate) applied on a 14-day schedule is recommended (3). Other fungicides are less effective.

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